HISTORICAL PERSPECTIVE



Cancer genes and the pathways they control

Bert Vogelstein & Kenneth W Kinzler

The revolution in cancer research can be summed up in a single sentence: cancer is, in essence, a genetic disease. In the last decade, many important genes responsible for the genesis of various cancers have been discovered, their mutations precisely identified, and the pathways through which they act characterized. The purposes of this review are to highlight examples of progress in these areas, indicate where knowledge is scarce and point out fertile grounds for future investigation.

What we know

The cast. Alterations in three types of genes are responsible for tumorigenesis: oncogenes, tumor-suppressor genes and stability genes (Tables 1 and 2). Unlike diseases such as cystic fibrosis or muscular dystrophy, wherein mutations in one gene can cause disease, no single gene defect 'causes' cancer. Mammalian cells have multiple safeguards to protect them against the potentially lethal effects of cancer gene mutations, and only when several genes are defective does an invasive cancer develop. Thus it is best to think of mutated cancer genes as contributing to, rather than causing, cancer.

Oncogenes are mutated in ways that render the gene constitutively active or active under conditions in which the wild-type gene is not. Oncogene activations can result from chromosomal translocations, from gene amplifications or from subtle intragenic mutations affecting crucial residues that regulate the activity of the gene product. For example, the most common activating mutation of BRAF in human cancers changes a valine to a glutamate at codon 599, a residue within the activation loop of the kinase domain¹. The activation loop is normally regulated by phosphorylation at adjacent residues (Thr598 and Ser601). This suggests that the glutamate substitution at codon 599 mimics a phosphate group and constitutively activates the enzyme even in the absence of signals that would normally result in phosphorylation of the adjacent threonine or serine residues. The activated BRAF kinase then phosphorylates downstream targets such as extracellular signal-regulated kinase (ERK), leading to aberrant growth²(Fig. 1). A mutation in an oncogene is analogous to a stuck accelerator in an automobile; the car still moves forward even when the driver removes his foot from it. An activating somatic mutation in one allele of an oncogene is generally sufficient to confer a selective growth advantage on the cell.

Tumor-suppressor genes are targeted in the opposite way by genetic alterations: mutations reduce the activity of the gene

The authors are at the Howard Hughes Medical Institute and The Sidney Kimmel Comprehensive Cancer Center, The Johns Hopkins University Medical Institutions, Baltimore, Maryland 21231, USA. e-mail: vogelbe@welch.jhu.edu or kinzlke@jhmi.edu

Published online 30 July 2004; doi:10.1038/nm1087

product. Such inactivations arise from missense mutations at residues that are essential for its activity, from mutations that result in a truncated protein, from deletions or insertions of various sizes, or from epigenetic silencing. A mutation in a tumor-suppressor gene is analogous to a dysfunctional brake in an automobile; the car doesn't stop even when the driver attempts to engage it. Some recently described tumor-suppressor genes have been hypothesized to exert a selective advantage on a cell when only one allele is inactivated and the other remains functional (that is, haploinsufficiency)³. However, mutations in both the maternal and paternal alleles of a tumor-suppressor gene are generally required to confer a selective advantage to the cell. This situation commonly arises through the deletion of one allele via a gross chromosomal event such as loss of an entire chromosome or chromosome armcoupled with an intragenic mutation of the other allele⁴.

Oncogene and tumor-suppressor gene mutations all operate similarly at the physiologic level: they drive the neoplastic process by increasing tumor cell number through the stimulation of cell birth or the inhibition of cell death or cell-cycle arrest. The increase can be caused by activating genes that drive the cell cycle, by inhibiting normal apoptotic processes or by facilitating the provision of nutrients through enhanced angiogenesis (Figs. 2-4). A third class of cancer genes, called stability genes or caretakers, promotes tumorigenesis in a completely different way when mutated. This class includes the mismatch repair (MMR), nucleotide-excision repair (NER) and base-excision repair (BER) genes responsible for repairing subtle mistakes made during normal DNA replication or induced by exposure to mutagens (Table 1). Other stability genes control processes involving large portions of chromosomes, such as those responsible for mitotic recombination and chromosomal segregation (for example, BRCA1, BLM and ATM; Table 1). Stability genes keep genetic alterations to a minimum, and thus when they are inactivated, mutations in other genes occur at a higher rate⁵. All genes are potentially affected by the resultant increased rate of mutation, but only mutations in oncogenes and tumor-suppressor genes affect net cell growth and can thereby confer a selective growth advantage to the mutant cell. As with tumor-suppressor genes, both alleles of stability genes generally must be inactivated for a physiologic effect to result.



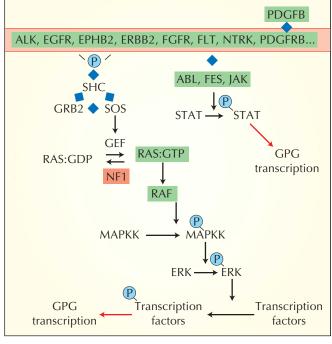


Figure 1 Receptor tyrosine kinase (RTK) pathway. Gene products in red boxes indicate that the corresponding genes are mutated in the germline (and generally also are somatically mutated in nonfamilial tumors; see Table 1). Gene products in green boxes indicate that the corresponding gene has been found to be mutated only somatically (see Table 2). 'GPG' denotes growth-promoting-genes—that is, genes that stimulate cell proliferation or inhibit the rate of cell death or arrest. Diamonds (♠) indicate protein-protein interactions. Red arrows and red T-bars indicate transcriptional induction and repression, respectively. Small circled 'P', 'OH' and 'Ub' represent covalently attached phosphate, hydroxyl and ubiquitin groups, respectively.

In the analogy to autos, stability genes represent the mechanics and a defective stability gene is akin to an inept mechanic.

Mutations in these three classes of genes can occur in the germline, resulting in hereditary predispositions to cancer (Table 1), or in single somatic cells, resulting in sporadic tumors (Tables 1 and 2). It is important to point out that a mutation is defined as any change in the sequence of the genome. These changes include those affecting single base pairs as well as those creating large or small deletions or insertions, amplifications or translocations. In the germline, the most common mutations are subtle (point mutations or small deletions or insertions), whereas all types of mutation can be found in tumor cells. In fact, cancers represent one of the few disease types in which somatic mutations occurring after birth are pathogenic.

The first somatic mutation in an oncogene or tumor-suppressor gene that causes a clonal expansion initiates the neoplastic process⁶. Subsequent somatic mutations result in additional rounds of clonal expansion (and thus in tumor progression)⁷. Indeed, the best modern definition of a neoplastic cell is one that has clonally expanded as a result of somatic mutations^{6,7}. Germline mutations of these genes cause cancer predisposition, not cancer *per se*: people with these mutations have a 'head start' on the neoplastic process, as a mutation that can contribute to cancer is already present in every one of their cells. Such individuals therefore often develop multiple tumors that occur at an earlier age than in individuals whose cancer-gene mutations have all occurred

somatically⁴. Examples of hereditary syndromes associated with inherited mutations are listed in Table 1. In people with these syndromes, only a very small fraction of the total cells in an at-risk organ become neoplastic because other (somatic) mutations are required to develop a clinically detectable lesion. In nearly all dominantly inherited syndromes caused by tumor-suppressor genes and stability genes, the first somatic mutation affects the normal copy of the gene inherited from the unaffected parent⁴. Interestingly, the most common forms of hereditary cancer predisposition, leading to breast and colon cancers, are caused by inherited mutations of stability genes rather than tumor-suppressor genes or oncogenes (Table 1).

The plot. As noted above, cancer-gene mutations enhance net cell growth. As a result of research performed over the past decade, it is clear that there are many fewer pathways than genes. This concept is very familiar to geneticists studying yeast, flies, mice or worms there are almost always a variety of genes that, when altered, lead to similar phenotypes^{8,9}. Application of this concept to cancer has been solidified by elucidation of the biochemical functions of the altered cancer genes, either in cell culture systems, in mice or in other organisms. For example, several cancer genes directly control transitions from a resting stage (G0 or G1) to a replicating phase (S) of the cell cycle (Fig. 2a, Rb pathway). The products of these genes include proteins as diverse as cdk4 (a kinase), cyclin D1 (which interacts with and activates cdk4), Rb (a transcription factor) and p16 (which interacts with and inhibits cdk4) 10-12. The genes encoding Rb and p16 are tumor-suppressor genes inactivated by mutation, whereas those encoding cdk4 and cyclin D1 are oncogenes activated by mutation. In addition to functional studies in model systems, detailed studies of individual tumor types have also provided compelling evidence that these four genes function in a single pathway in human cancers. Such studies have shown that the mutations within this pathway obey an 'exclusivity principle': that is, one and only one of the four genes noted above is generally mutated in any single tumor, exactly as predicted if the functional effect of each mutation was similar^{10–13}.

Another example of the reason for focusing on pathways rather than individual genes has been provided by studies of the *TP53* tumor-suppressor gene. The p53 protein is a transcription factor that normally inhibits cell growth and stimulates cell death when induced by cellular stress^{14–16}. The most common way to disrupt the p53 pathway is through a point mutation that inactivates its capacity to bind specifically to its cognate recognition sequence. However, there are several other ways to achieve the same effect, including amplification of the *MDM2* gene and infection with DNA tumor viruses whose products (such as the E6 protein of human papilloma virus) bind to p53 and functionally inactivate it (Fig. 2b, p53 pathway).

One of the most important—and most curious—discoveries of the 1990s was that virtually all DNA tumor viruses that cause tumors in experimental animals or humans encode proteins that inactivate both Rb and p53 (refs. 17–19). Of the hundreds of cancer genes known or remaining to be discovered, why should these two have been singled out as targets for inactivation by all DNA tumor viruses? The answer may be that it is impossible for a tumor of epithelial origin to form unless the p53 and Rb tumor-suppressor gene pathways have been inactivated. This conjecture is supported by studies showing that these two pathways are altered in a large fraction of many types of cancers. We predict that most of the cancers that now appear to be devoid of mutations in these two



Table 1 Cancer predisposition genes

FAP Dominant Inactivation of WT allele APC Colon, thyroid, stomach, intestin MXM2 Attenuated polyposis Dominant Inactivation of WT allele APC Stomach Stom	Gene (synonym(s)) ^a	Syndrome	Hereditary pattern	Second hit	Pathway ^b	Major heredity tumor types ^c
Author Authorized polyposis Dominant Inactivation of WT allele APC Stomach Familial gastric carcinome Dominant Inactivation of WT allele APC Stomach APC A	Tumor-suppressor genes					
Canderin Familial gastric carcinoma Dominant Inactivation of WT allele APC Stomach Simpson Galbi-Belmela AVI	APC					
Simpson-Golabi-Belmel syndrome syndrome syndrome syndrome portional syndromes should be syndrome portional syndromes should be syndrome portional syndromes should be						
Syndrome XT1.2	,	9				
Process Proc	GPC3	Simpson-Golabi-Behmel	X-linked	?	APC	Embryonal
Description		syndrome				
exostoses SUFU Medulloblastoma Dominant Inactivation of WT allele GLI Skin, medulloblastoma predisposition Fig. Hereditary lelomymatosis Dominant Inactivation of WT allele HIFI Lelomymans predisposition Familial paraganglioma Dominant Inactivation of WT allele HIFI Lelomymans Fig. 10 No. 1	CYLD	Familial cylindromatosis	Dominant	Inactivation of WT allele	APOP	Pilotrichomas
### PATCH	EXT1,2	Hereditary multiple	Dominant	Inactivation of WT allele	GLI	Bone
Mediloblastoma predisposition Inactivation of WT allele GLI Skin, meduloblastoma predisposition Femilial paragangloma Dominant Inactivation of WT allele HIF1 Leiomyomas Paragangloma Dominant Inactivation of WT allele HIF1 Paraganglomas Phecitomocytomas Phecitomocyto		exostoses				
predisposition H Hereditary leiomyomatosis Dominant Inactivation of WT allele HIF1 Paraganglomas, phoeobromocytomas phoeobromocytomas phoeobromocytomas (Aidney PS3 (pS3)) HL Von Hippel-Lindau syndrome Dominant Inactivation of WT allele HIF1 Kidney PS3 (pS3) Li-Fraumeni syndrome Dominant Inactivation of WT allele p53 Breast, sarcoma, adrenal, brain WTI Familial Wilms tumor Dominant Inactivation of WT allele p53 Wilms' STATI (LKRI) Peutz-leghers syndrome Dominant Inactivation of WT allele P13K Hamartoma, glioma, uterus PTR (Cowden syndrome Dominant Inactivation of WT allele P13K Hamartoma, glioma, uterus P18KI, p18WI, p18WI	PTCH	Gorlin syndrome	Dominant	Inactivation of WT allele	GLI	Skin, medulloblastoma
Hereditary leiomyomatosis Dominant Inactivation of WT allele HIFI Leiomyomas	SUFU	Medulloblastoma	Dominant	Inactivation of WT allele	GLI	Skin, medulloblastoma
Paraganglomes Parillal paraganglomes Par		predisposition				
Paraganglomes Parillal paraganglomes Par	FH	Hereditary leiomyomatosis	Dominant	Inactivation of WT allele	HIF1	Leiomyomas
### Photochromecytomas #### Photochromecytomas ##### Photochromecytomas ####################################						*
### VPT Ven Hippel-Lindau syndrome Dominant Inactivation of WT allele P53 Breast, sarcoma, adrenal, brain ### VPT Familial Wilms tumor Dominant Inactivation of WT allele P53 Wilms ### VPT Familial Wilms tumor Dominant Inactivation of WT allele P53 Wilms ### VPT Cowden syndrome Dominant Inactivation of WT allele P13K Hamardona, gloma, uterus ### VPT Cowden syndrome Dominant Inactivation of WT allele P13K Hamardona, gloma, uterus ### VPT Cowden syndrome Dominant Inactivation of WT allele P13K Hamardona, gloma, uterus ### VPT Cowden syndrome Dominant Inactivation of WT allele P13K Hamardona, gloma, uterus ### VPT Pamilial malignant Dominant Inactivation of WT allele P13K Hamardona, gloma, uterus ### VPT Pamilial malignant Dominant Inactivation of WT allele P13K Hamardona, gloma, uterus ### VPT Pamilial malignant Dominant Inactivation of WT allele P13K Melanoma ### Pamilial malignant Dominant Inactivation of WT allele RB Eye ### WELL Neurofibromatosis type 1 Dominant Inactivation of WT allele RB Eye ### VPT Neurofibromatosis type 1 Dominant Inactivation of WT allele SMAD Gastrointestinal ### VPT Juvenile polyposis Dominant Inactivation of WT allele SMAD Gastrointestinal ### Pamilial malignant Dominant Inactivation of WT allele SMAD Parathyroid, pituitary, islet cell, carcinoid ### Complex SMAD Satrointestinal SMAD Parathyroid, jawr fibroma ### Pamilial malignant Dominant Inactivation of WT allele P15K Parathyroid, jawr fibroma ### Pamilial malignant Dominant Inactivation of WT allele P15K Parathyroid, jawr fibroma ### Pamilial malignant Dominant Inactivation of WT allele P15K Parathyroid, jawr fibroma ### Pamilial malignant Dominant Inactivation of WT allele P15K Parathyroid, jawr fibroma ### Pamilial malignant Dominant Inactivation of WT allele P15K Parathyroid, j	00110, 0, 0	r anniar paragangnoma	Dominum	mactivation of W1 ancie	1111 1	
PFS3 (pS3)	VHI	Von Hinnel I indau syndrome	Dominant	Inactivation of WT allele	HIF1	-
WT1 Familial Wilms tumor Dominant Inactivation of WT allele p53 Wilms' 5TK11 (LKB1) Peutz-leghers syndrome Dominant Inactivation of WT allele P13K Intestinal, ovarian, pancreatic PERN Cowden syndrome Dominant Inactivation of WT allele P13K Hamartoma, gliome, uterus DOM/A2A Familial malignant Dominant Inactivation of WT allele RB Melanoma DDM4 Familial malignant Dominant Inactivation of WT allele RB Melanoma RB1 Hereditary retinoblastoma Dominant Inactivation of WT allele RB Eye WF1 Neurofibromatosis type 1 Dominant Inactivation of WT allele RTK Neurofibroma MBMP1A Juvenile polyposis Dominant Inactivation of WT allele SMAD Gastrointestinal MBMP1A Juvenile polyposis Dominant Inactivation of WT allele SMAD Parathyroid, pituitary, islet cell, carcinoid MBMP1A Juvenile polyposis Dominant Inactivation of WT allele SMAD Parathyroid, pituitary,						
### Familial Wilms tumor ### Dominant ### Domin	1P53 (p53)	Li-Fraumeni syndrome	Dominant	inactivation of WT allele	p53	
Particular Pentral P						
Dominant Inactivation of WT allele PI3K Hamartoma, glioma, uterus	WT1	Familial Wilms tumor	Dominant	Inactivation of WT allele	p53	
Figure F	STK11 (LKB1)	Peutz-Jeghers syndrome	Dominant	Inactivation of WT allele	PI3K	Intestinal, ovarian, pancreatic
DOMINA Pamilial malignant melanoma Pamilial melanoma Pamilial malignant melanoma Pamil	PTEN	Cowden syndrome	Dominant	Inactivation of WT allele	PI3K	Hamartoma, glioma, uterus
melanoma pl.f6 ^{INK4A} , p.14 ^{MRT}) melanoma pl.f6	TSC1, TSC2	Tuberous sclerosis	Dominant	Inactivation of WT allele	PI3K	Hamartoma, kidney
melanoma pl.f6 ^{INK4A} , p.14 ^{MRT}) melanoma pl.f6	CDKN2A	Familial malignant	Dominant	Inactivation of WT allele	RR	
Pamilial malignant melanoma		•	Dominant	maceración de tre anoid		moranoma, panereae
Membra M			Dominant	?	RB	Melanoma
Hereditary retinoblastoma Dominant Inactivation of WT allele RTK Neurofibromatosis type 1 Dominant Inactivation of WT allele RTK Neurofibroma		•	Dominant	•		
Neurofibromatosis type 1 Dominant Inactivation of WT allele RTK Neurofibroma New Parathyroid, pituitary, islet cell, neoplasia type 1 Neurofibromatosis type 2 Neurofibromatosis type 4 Neurofibroma	RR1		Dominant	Inactivation of WT allele	RB	Eve
Juvenile polyposis Dominant Inactivation of WT allele SMAD Parathyroid, pituitary, islet cell, carcinoid neoplasia type I puvenile polyposis Dominant Inactivation of WT allele SMAD Gastrointestinal Parathyroid, pituitary, islet cell, carcinoid Gastrointestinal Parathyroid pituitary, islet cell, carcinoid Gastrointestinal Parathyroid, jaw fibroma Jaw-tumor syndrome. WF2 Neurofibromatosis type 2 Dominant Inactivation of WT allele ? Renal, hair follicle Parathyroid, jaw fibroma Jaw-tumor syndrome. WF2 Neurofibromatosis type 2 Dominant Inactivation of WT allele ? Meningioma, acoustic neuroma Stability genes WUTHH Attenuated polyposis Recessive ? BER Colon ATM Attenuated polyposis Recessive ? CIN Leukemias, lymphomas, brain Bloom syndrome Recessive ? CIN Leukemias, lymphomas, skin Bloom syndrome Recessive ? CIN Leukemias, lymphomas, skin Bloom syndrome Recessive ? CIN Leukemias Parathyroid, parathyroid, parathyroid, acrenal tumors stromal tumors WRS1 Nijmegen breakage syndrome Recessive ? CIN Lymphomas, brain Recessive Parathyroid, parathyroid, adrenal tumors stromal tumors WRF2 Neuror spidnent Parathyroid Parathyroid, adrenal tumors stromal tumors WET Hereditary papillary renal Dominant Parathyroid, adrenal tumors stromal tumors WET Hereditary papillary renal Dominant Parathyroid, adrenal tumors stromal tumors WET Multiple endocrine Dominant Mutant allele duplication RTK Thyroid, parathyroid, adrenal						
Multiple endocrine neoplasia type I Dominant neoplasia type I Dominant neoplasia type I Juvenile polyposis Dominant Inactivation of WT allele SMAD Gastrointestinal BMAD Birt-Hogg-Dube syndrome Hyperparathyroidism Dominant Inactivation of WT allele Practivation of WT allele Pra						
neoplasia type I Juvenile polyposis Dominant Inactivation of WT allele SMAD Gastrointestinal SMAD4 (DPC4) Juvenile polyposis Dominant Inactivation of WT allele ? Renal, hair follicle Parathyroid; jaw fibroma Jaw-tumor syndrome. WF2 Neurofibromatosis type 2 Dominant Inactivation of WT allele ? Parathyroid, jaw fibroma Jaw-tumor syndrome. WF2 Neurofibromatosis type 2 Dominant Inactivation of WT allele ? Meningioma, acoustic neuroma Stability genes WUTYH Attenuated polyposis Recessive ? BER Colon WTM Ataxia telangiectasia Recessive ? CIN Leukemias, lymphomas, brain Bloom syndrome Recessive ? CIN Leukemias, lymphomas, brain Inactivation of WT allele CIN Breast, ovary ARNCA, C, D2, E, F,G Fanconi anemia Recessive ? CIN Leukemias WBS1 Nijmegen breakage syndrome Recessive ? CIN Lymphomas, brain RecCQL4 Rothmund-Thomson syndrome Recessive ? CIN Bone, skin Bone, skin Bone, skin Bone, skin Recessive ? CIN Bone, skin Bone, brain Bone,			Dominant	Inactivation of WT allele		Gastrointestinal
SMAD Gastrointestinal SMAD Gastrointestinal SMAD Gastrointestinal SMAD	MEN1	Multiple endocrine	Dominant	Inactivation of WT allele	SMAD	Parathyroid, pituitary, islet cell,
Birt-Hogg-Dube syndrome Dominant Inactivation of WT allele ? Renal, hair follicle Hyperparathyroidism Dominant Jaw-tumor syndrome. WF2 Neurofibromatosis type 2 Dominant Inactivation of WT allele ? Parathyroid, jaw fibroma Jaw-tumor syndrome. WF2 Neurofibromatosis type 2 Dominant Inactivation of WT allele ? Meningioma, acoustic neuroma stability genes WUTH Attenuated polyposis Recessive ? BER Colon ATM Ataxia telangiectasia Recessive ? CIN Leukemias, lymphomas, brain Recessive Prancial Recessive Pra		neoplasia type I				carcinoid
Hyperparathyroidism Jaw-tumor syndrome. WF2 Neurofibromatosis type 2 Dominant Inactivation of WT allele ? Meningioma, acoustic neuroma stability genes MUTYH Attanuated polyposis Recessive ? BER Colon ATM Ataxia telangiectasia Recessive ? CIN Leukemias, lymphomas, brain Blom syndrome Recessive ? CIN Leukemias, lymphomas, skin Blom syndrome Recessive ? CIN Leukemias, lymphomas, skin Blom syndrome Recessive ? CIN Leukemias, lymphomas, skin BRCA1, BRCA2 Hereditary breast cancer Dominant Inactivation of WT allele CIN Breast, ovary CIN Leukemias AND Nijmegen breakage syndrome Recessive ? CIN Leukemias AND Nijmegen breakage syndrome Recessive ? CIN Bone, skin Bone, skin Recessive Recessive ? CIN Bone, skin Bone, skin Bone, brain AND Nijmegen breakage syndrome Recessive ? CIN Bone, skin Bone, skin Bone, brain AND Nijmegen breakage syndrome Recessive ? CIN Bone, skin Bone, skin Bone, skin Bone, brain AND BONE, skin Bone, brain AND BONE, brain	SMAD4 (DPC4)	Juvenile polyposis	Dominant	Inactivation of WT allele	SMAD	Gastrointestinal
Jaw-tumor syndrome. Neurofibromatosis type 2 Dominant Inactivation of WT allele ? Meningioma, acoustic neuroma Stability genes WUTYH Attenuated polyposis Recessive ? BER Colon ATM Ataxia telangiectasia Recessive ? CIN Leukemias, lymphomas, brain BIM BIDOM syndrome Recessive ? CIN Leukemias, lymphomas, skin BIRCA1, BRCA2 Hereditary breast cancer Dominant Inactivation of WT allele CIN Breast, ovary Fanconi anemia Recessive ? CIN Leukemias WBS1 Nijmegen breakage syndrome Recessive ? RECGU4 Rothmund-Thomson syndrome Recessive ? RECSUA ROTHMUND-THOMSON Syndrome Recessive ? RECSUA ROTHMUND-THOMSON Syndrome Recessive ? WBN Werner syndrome Recessive ? CIN Bone, skin WSH2, MLH1, HNPCC Dominant Inactivation of WT allele MMR Colon, uterus WSH2, MLH1, HNPCC Dominant Inactivation of WT allele MMR Colon, uterus WSH3, MSH4, MLH1, HNPCC Dominant Recessive ? RECESSIVE ? NER Skin DOM2 Domicogenes KIT Familial gastrointestinal Stromal tumors Stromal tumors WET Hereditary papillary renal Dominant ? RECH Hereditary papillary renal Dominant ? RECH ROTHMUND-S MUTYH Attenuated polyposis Recessive ? RTK Gastrointestinal stromal tumors STRIM Gastrointestinal stromal tumors RTK Kidney CPDGFRA Familial gastrointestinal Dominant ? RTK Gastrointestinal stromal tumors STRIM Gastrointestinal stromal tumors Mutant allele duplication RTK Thyroid, parathyroid, adrenal	BHD	Birt-Hogg-Dube syndrome	Dominant	Inactivation of WT allele	?	Renal, hair follicle
Neurofibromatosis type 2 Dominant Inactivation of WT allele ? Meningioma, acoustic neuroma Rability genes MUTYH Attenuated polyposis Recessive ? BER Colon NaTM Ataxia telangiectasia Recessive ? CIN Leukemias, lymphomas, brain Bloom syndrome Recessive ? CIN Leukemias, lymphomas, brain BRCA1, BRCA2 Hereditary breast cancer Dominant Inactivation of WT allele CIN Breast, ovary CIN Leukemias NBS1 Nijmegen breakage syndrome Recessive ? CIN Lymphomas, brain RECQL4 Rothmund-Thomson syndrome Recessive ? CIN Lymphomas, brain BRCQL4 Rothmund-Thomson syndrome Recessive ? CIN Bone, skin WRNN Werner syndrome Recessive ? CIN Bone, brain NBS1 NIJMEGEN BRCA2 ROTHMUND-THOMSON SYNDROME RECESSIVE ? CIN Bone, brain NBCQL4 ROTHMUND-THOMSON SYNDROME RECESSIVE ? CIN Bone, brain NBCQCL5 RECC2-5; Xeroderma pigmentosum Recessive ? NER Skin NBCQCDB2 NDCOGENES NET Familial gastrointestinal Stromal tumors stromal tumors NBCC RECESSIVE ? RTK Kidney CEDGFRA Familial gastrointestinal Dominant ? RTK Kidney RECESSIVE RTK Gastrointestinal stromal tumors stromal tumors NBCC RTK Gastrointestinal stromal tumors NBCC RTK Thyroid, parathyroid, adrenal	HRPT2	Hyperparathyroidism	Dominant	Inactivation of WT allele	?	Parathyroid, jaw fibroma
Stability genes MUTYH Attenuated polyposis Recessive ? BER Colon ATM Ataxia telangiectasia Recessive ? CIN Leukemias, lymphomas, brain Bloom syndrome Recessive ? CIN Leukemias, lymphomas, skin Bloom syndrome Recessive ? CIN Leukemias, lymphomas, skin BRCA1, BRCA2 Hereditary breast cancer Dominant Inactivation of WT allele CIN Breast, ovary FANCA, C, D2, E, F,G Fanconi anemia Recessive ? CIN Leukemias WBS1 Nijmegen breakage syndrome Recessive ? RECQL4 Rothmund-Thomson syndrome Recessive ? WBRN Werner syndrome Recessive ? WBRN WERN Werner syndrome Recessive ? WBRN WERN WERNER WBRN WERNER WBRN WERN WERNER WBRN WERNER WB		Jaw-tumor syndrome.				
Recessive ? BER Colon Attanuated polyposis Recessive ? CIN Leukemias, lymphomas, brain Blom syndrome Recessive ? CIN Leukemias, lymphomas, brain Blom syndrome Recessive ? CIN Leukemias, lymphomas, skin Blom syndrome Recessive ? CIN Leukemias, lymphomas, skin BRCA1, BRCA2 Hereditary breast cancer Dominant Inactivation of WT allele CIN Breast, ovary FANCA, C, D2, E, F,G Fanconi anemia Recessive ? CIN Leukemias RECQL4 Rothmund-Thomson syndrome Recessive ? CIN Bone, skin WHRN Werner syndrome Recessive ? CIN Bone, brain WHRD Colon, uterus WHRD Colon, uterus WHRD Colon, uterus WHRD Colon, uterus WHRD RECESSIVE ? RECESSIVE ? RECRUPTED RECESSIVE RECESSIVE RECESSIVE RECESSIVE ? RECESSIVE RECESS	NF2	Neurofibromatosis type 2	Dominant	Inactivation of WT allele	?	Meningioma, acoustic neuroma
MUTYH Attenuated polyposis Recessive ? BER Colon ATM Ataxia telangiectasia Recessive ? CIN Leukemias, lymphomas, brain Bloom syndrome Recessive ? CIN Leukemias, lymphomas, brain Bloom syndrome Recessive ? CIN Leukemias, lymphomas, skin BRCA1, BRCA2 Hereditary breast cancer Dominant Inactivation of WT allele CIN Breast, ovary RANCA, C, D2, E, F,G Fanconi anemia Recessive ? CIN Leukemias WBS1 Nijmegen breakage syndrome Recessive ? CIN Lymphomas, brain RECQL4 Rothmund-Thomson syndrome Recessive ? CIN Bone, skin Bene, skin RECQL4 Rothmund-Thomson syndrome Recessive ? CIN Bone, skin Bone, brain Inactivation of WT allele MMR Colon, uterus WBH2, MLH1, HNPCC Dominant Inactivation of WT allele MMR Colon, uterus WBH2, CFA, C; ERCC2-5; Xeroderma pigmentosum Recessive ? NER Skin WBCDDB2 WICH Familial gastrointestinal Dominant ? RTK Gastrointestinal stromal tumors stromal tumors WBET Hereditary papillary renal Cell carcinoma PDGFRA Familial gastrointestinal Dominant ? RTK Gastrointestinal stromal tumors stromal tumors WET Hereditary papillary renal Dominant ? RTK Gastrointestinal stromal tumors stromal tumors WET Multiple endocrine Dominant Mutant allele duplication RTK Thyroid, parathyroid, adrenal	Stahility genes					
Ataxia telangiectasia Recessive ? CIN Leukemias, lymphomas, brain Bloom syndrome Recessive ? CIN Leukemias, lymphomas, skin BRCA1, BRCA2 Hereditary breast cancer Dominant Inactivation of WT allele CIN Breast, ovary CIN Leukemias PANCA, C, D2, E, F, G Fanconi anemia Recessive ? CIN Lymphomas, brain RECQL4 Rothmund-Thomson syndrome Recessive ? CIN Lymphomas, brain RECQL4 Rothmund-Thomson syndrome Recessive ? CIN Bone, skin Bone, skin Recessive ? CIN Bone, brain RECQL4 Rothmund-Thomson syndrome Recessive ? Recessive Recessive ? Recessive R		Attenuated polyposis	Recessive	?	BFR	Colon
Blom syndrome Recessive ? CIN Leukemias, lymphomas, skin BRCA1, BRCA2 Hereditary breast cancer Dominant Inactivation of WT allele CIN Breast, ovary CIN Leukemias WBS1 Nijmegen breakage syndrome Recessive ? CIN Lymphomas, brain RECQL4 Rothmund-Thomson syndrome Recessive ? CIN Bone, skin Bone, skin Bone, skin Bone, brain CIN Bone, br						
Hereditary breast cancer Pominant Recessive ? CIN Leukemias Nijmegen breakage syndrome Recessive ? CIN Lymphomas, brain ReCQL4 Rothmund-Thomson syndrome Recessive ? CIN Bone, skin Bone, skin Bone, skin Recessive ? CIN Bone, brain RecQL4 Rothmund-Thomson syndrome Recessive ? CIN Bone, brain Recessive Praint Recessive Recessive Praint Recessive		•		•		
Recessive ? CIN Leukemias Nijmegen breakage syndrome Recessive ? CIN Lymphomas, brain RECQL4 Rothmund-Thomson syndrome Recessive ? CIN Bone, skin WRN Werner syndrome Recessive ? CIN Bone, brain MSH2, MLH1, HNPCC Dominant Inactivation of WT allele MMR Colon, uterus WRHA, C; ERCC2-5; Xeroderma pigmentosum Recessive ? NER Skin DOB2 Nocogenes K(IT Familial gastrointestinal Stromal tumors stromal tumors ACT Hereditary papillary renal Cell carcinoma PDGFRA Familial gastrointestinal Dominant ? RTK Gastrointestinal stromal tumors stromal tumors MUTH Hereditary papillary renal Dominant ? RTK Gastrointestinal stromal tumors stromal tumors MUTH Hereditary papillary renal Dominant ? RTK Gastrointestinal stromal tumors stromal tumors MUTH Hereditary papillary renal Dominant ? RTK Gastrointestinal stromal tumors stromal tumors MUTH Hereditary papillary renal Dominant ? RTK Gastrointestinal stromal tumors MUTH Hereditary papillary renal Dominant ? RTK Gastrointestinal stromal tumors MUTH Hereditary papillary renal Dominant ? RTK Gastrointestinal stromal tumors MUTH Hereditary papillary renal Dominant ? RTK Gastrointestinal stromal tumors MUTH HEREDITARY RECESSIVE ? CIN Lymphomas, brain MUTH HOND MITH HO		-		•		
NBS1 Nijmegen breakage syndrome Recessive ? CIN Lymphomas, brain RECQL4 Rothmund-Thomson syndrome Recessive ? CIN Bone, skin Bone, skin WRN Werner syndrome Recessive ? CIN Bone, brain CIN Bone, brain MSH2, MLH1, HNPCC Dominant Inactivation of WT allele MMR Colon, uterus MSH6, PMS2 PobB2 NCPA, C; ERCC2–5; Xeroderma pigmentosum Recessive ? NER Skin MCDB2 Nacogenes (IT Familial gastrointestinal stromal tumors stromal tumors MET Hereditary papillary renal Dominant Perditary papillary renal cell carcinoma PobB2 PDGFRA Familial gastrointestinal Dominant ? RTK Gastrointestinal stromal tumors stromal tumors MET Hereditary papillary renal Dominant Pominant Pominant RTK Gastrointestinal stromal tumors stromal tumors MUT Hereditary papillary renal Dominant Pominant RTK Gastrointestinal stromal tumors stromal tumors MUT Hereditary papillary renal Dominant RTK Gastrointestinal stromal tumors stromal tumors		•				
RECQL4 Rothmund-Thomson syndrome Recessive Rec	ranua, u, U∠, E, t,G	гансоні апетна	Recessive	:	CIN	LeukeiiiiaS
RECQL4 Rothmund-Thomson syndrome Recessive Rec	NDC1	Niimaanan haaal	Danasia	2	OIN	Luman barrana barra
WRN Werner syndrome Recessive ? CIN Bone, brain MSH2, MLH1, HNPCC Dominant Inactivation of WT allele MMR Colon, uterus WSH6, PMS2 WPA, C; ERCC2-5; Xeroderma pigmentosum Recessive ? NER Skin DOB2 Docogenes WIT Familial gastrointestinal Dominant ? RTK Gastrointestinal stromal tumors stromal tumors MET Hereditary papillary renal Dominant Mutant allele duplication RTK Kidney CPDGFRA Familial gastrointestinal Stromal tumors STET Multiple endocrine Dominant Mutant allele duplication RTK Thyroid, parathyroid, adrenal						
MSH2, MLH1, HNPCC Dominant Inactivation of WT allele MMR Colon, uterus (PA, C; ERCC2-5; Xeroderma pigmentosum Recessive ? NER Skin (PDB2 (MT Familial gastrointestinal pominant ? RTK Gastrointestinal stromal tumors stromal tumors (MET Hereditary papillary renal pominant Mutant allele duplication RTK Kidney cell carcinoma (PDGFRA Familial gastrointestinal pominant ? RTK Gastrointestinal stromal tumors stromal tumors (MET Hereditary papillary renal pominant ? RTK Gastrointestinal stromal tumors (PDGFRA Familial gastrointestinal pominant ? RTK Gastrointestinal stromal tumors (RET Multiple endocrine Dominant Mutant allele duplication RTK Thyroid, parathyroid, adrenal	· · ·	*		•		
MSH6, PMS2 APA, C; ERCC2–5; Xeroderma pigmentosum Recessive ? NER Skin DDB2 Dincogenes KIT Familial gastrointestinal Dominant ? RTK Gastrointestinal stromal tumors stromal tumors MET Hereditary papillary renal Dominant Mutant allele duplication RTK Kidney cell carcinoma PDGFRA Familial gastrointestinal Dominant ? RTK Gastrointestinal stromal tumors stromal tumors RET Multiple endocrine Dominant Mutant allele duplication RTK Thyroid, parathyroid, adrenal				·		
Concogenes Concog	MSH2, MLH1, MSH6, PMS2	HNPCC	Dominant	Inactivation of WT allele	MMR	Colon, uterus
Concogenes K(IT Familial gastrointestinal Dominant ? RTK Gastrointestinal stromal tumors stromal tumors MET Hereditary papillary renal Dominant Mutant allele duplication RTK Kidney cell carcinoma PDGFRA Familial gastrointestinal Dominant ? RTK Gastrointestinal stromal tumors stromal tumors RET Multiple endocrine Dominant Mutant allele duplication RTK Thyroid, parathyroid, adrenal	XPA, C; ERCC2-5;	Xeroderma pigmentosum	Recessive	?	NER	Skin
Familial gastrointestinal Dominant ? RTK Gastrointestinal stromal tumors stromal tumors MET Hereditary papillary renal Dominant Mutant allele duplication RTK Kidney cell carcinoma PDGFRA Familial gastrointestinal Dominant ? RTK Gastrointestinal stromal tumors stromal tumors RET Multiple endocrine Dominant Mutant allele duplication RTK Thyroid, parathyroid, adrenal						
stromal tumors MET Hereditary papillary renal Dominant Mutant allele duplication RTK Kidney cell carcinoma PDGFRA Familial gastrointestinal Dominant ? RTK Gastrointestinal stromal tumors stromal tumors RET Multiple endocrine Dominant Mutant allele duplication RTK Thyroid, parathyroid, adrenal	-	Esmilial gastraintestinal	Dominant	2	DTV	Contraintactinal attendal turns
Hereditary papillary renal Dominant Mutant allele duplication RTK Kidney cell carcinoma PDGFRA Familial gastrointestinal Dominant ? RTK Gastrointestinal stromal tumors stromal tumors RET Multiple endocrine Dominant Mutant allele duplication RTK Thyroid, parathyroid, adrenal	NI I	_	זווווווסעד	:	KIK	Gastrointestinai stromai tumors
cell carcinoma PDGFRA Familial gastrointestinal Dominant ? RTK Gastrointestinal stromal tumors stromal tumors RET Multiple endocrine Dominant Mutant allele duplication RTK Thyroid, parathyroid, adrenal	MET		Danimant	Modern allele College	DTI	V:
PDGFRA Familial gastrointestinal Dominant ? RTK Gastrointestinal stromal tumors stromal tumors RET Multiple endocrine Dominant Mutant allele duplication RTK Thyroid, parathyroid, adrenal	MET		Dominant	wutant allele duplication	RIK	Kidney
stromal tumors RET Multiple endocrine Dominant Mutant allele duplication RTK Thyroid, parathyroid, adrenal	222524				DTI	
RET Multiple endocrine Dominant Mutant allele duplication RTK Thyroid, parathyroid, adrenal	PUGFKA	-	Dominant	<i>f</i>	RIK	Gastrointestinal stromal tumors
	0.57				D.T.	
neoplasia type II	RET		Dominant	Mutant allele duplication	RIK	Thyroid, parathyroid, adrenal
		neoplasia type II				

WT, wild type. ^aRepresentative genes of all the major pathways and hereditary cancer predisposition types are listed. For a complete list, see ref. 117. Approved gene symbols are provided for each entry, with alternative names in parentheses. ^bIn many cases, the gene has been implicated in several pathways. The single pathway that is listed for each gene represents a 'best guess' (when one can be made) and for the reasons noted in the text and in the legend to **Figure 9**, should not be regarded as conclusive. APOP, apoptotic pathway; RTK, receptor tyrosine kinase pathway (see **Fig. 1**). ^cIn most cases, the nonfamilial tumor spectrum caused by somatic mutations of the gene includes those occurring in the familial cases plus additional tumor types. For example, mutations of *TP53* and *CDKN2A* are found in many more tumor types than those to which Li-Fraumeni and familial malignant melanoma patients, respectively, are predisposed.



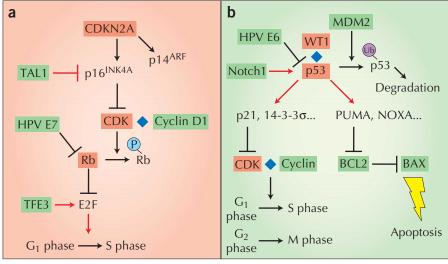


Figure 2 Rb (a) and p53 (b) pathways. Symbols as in Figure 1.

pathways will eventually be shown to contain them. This prediction will become testable once more efficient methods of detecting mutations are developed and all the genes in the pathways become known.

In addition to the Rb and p53 pathways, there are others that have a role in many tumor types, including those involving adenomatous polyposis coil (APC), glioma-associated oncogene (GLI), hypoxia-inducible transcription factor (HIF)-1, phosphoinositide 3-kinase (PI3K), SMADs and receptor tyrosine kinases (RTKs) (Figs. 1 and 4–8). In each case, mutations in multiple members of the pathway have been found to be mutated in more than one type of cancer, and in many cases the mutations within a single pathway obey the exclusivity principle noted above. The discoveries of these signal transduction pathways represent major scientific success stories of the last decade²⁰. Their delineation has practical as well as

theoretical implications. For example, they predict that targeted therapies directed against a particular gene product may be active against a tumor with a mutation of the targeted gene or any gene upstream of the target. Additionally, the fact that defects in a relatively small number of pathways underlie many different tumor types suggests that targeted therapeutics will be effective against a broad range of cancers.

Tumors can be broadly classified as liquid or solid. The former includes leukemias and lymphomas, composed of neoplastic cells whose precursors are normally mobile. Solid tumors are composed of epithelial or mesenchymal cells that normally are immobile. There are numerous other differences between liquid and solid tumors. For example, at least three mutations seem to be required to develop a malignant solid tumor in adults²¹; each of these mutations likely alters one of the pathways described in Figure 9. In contrast, only one or two mutations may be required to develop a malignant liquid tumor⁶. Perhaps liquid tumors don't require as many pathways to be inactivated because their precursor cells are already mobile and invasive, key characteristics that solid tumor cells must develop to become malignant. Additionally, oncogene activations caused by chromosome translocation events are the most common genetic alterations observed in liquid tumors²². Consistent chromosome translocations are much less common in solid tumors, whereas inactivations of tumor-suppressor genes are ubiquitious²³. Finally, there are several genes that are uniquely altered in specific subtypes of liquid tumors, seem primarily to affect differentiation, do not obviously participate in the pathways depicted in Figure 9 and do not occur in hereditary form (Table 2). These molecular distinctions add to the cytogenetic, epidemiologic and medical evidence that liquid and solid tumors should be considered separately in

terms of their biology, behavior and pathogenesis. As solid tumors are much more common than liquid tumors, most of the examples provided in this review focus on the former.

Supporting cast. That solid tumors are composed of two compartments, one consisting of neoplastic epithelial cells and the other of stromal cells, was pointed out a hundred years ago²⁴. The importance of the interactions between stroma and epithelium is becoming increasingly recognized^{25,26}. And four discoveries made during the last ten years have propelled one component of the stroma, endothelial cells, into the spotlight.

First, naturally occurring inhibitors of angiogenesis were identified and shown to hinder the growth of experimental tumors^{27,28}. Second, synthetic inhibitors of one of the major regulatory pathways of angiogenesis (VEGF) were produced and shown to inhibit

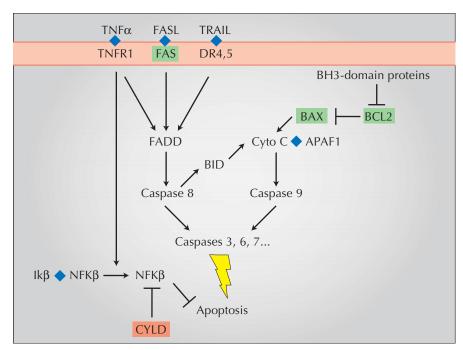


Figure 3 Apoptosis pathway. Symbols as in Figure 1.



tumor progression in patients with cancer²⁹. Third, inactivation of VHL, a classic tumor-suppressor gene, was shown to support renal cell tumor growth through the control of tumor angiogenesis³⁰. The protein encoded by VHL is part of a ubiquitin ligase complex that degrades HIF-1 α in the presence of oxygen (Fig. 4, HIF1 pathway). In the absence of oxygen under normal circumstances or when VHL is mutated in tumors, the HIF-1α transcription factor is stabilized, leading to the expression of cytokines such VEGF and culminating in angiogenesis31. And finally, all oncogenes and tumor-suppressor gene pathways have been implicated in angiogenesis, either directly or indirectly, as illustrated by the central position of the HIF1 pathway in Figure 9. An understanding of tumor angiogenesis, blood flow, oxygenation and related issues involving the tumor-host relationship is becoming essential to studies of cancer biology as well as to the design of more effective forms of cancer therapy.

What we don't know

The opening act. The process of tumorigenesis is initiated when a replication-competent cell (stem cell or partially differentiated descendent of a stem cell) acquires a mutation in a 'gatekeeping' pathway that endows it with a selective growth advantage. In some cancers, the gatekeeper has been identified (examples are RB1, APC and NF1 in tumors of the eye, colon and nervous system, respectively). In most common tumors, however, the gatekeeper is not known. It is also not known whether cancers of the lung, breast, prostate, bladder or brain can each be initiated through any one of several gatekeeping pathways or through only one. Many of the known gatekeepers were identified through the study of unusual families with predispositions to specific types of cancers⁴. There may be other families that provide clues to the nature of the gatekeepers in the future. We predict, however, that many of these remaining gatekeeping genes will only be identified through more 'brute-force' approaches involving sequence determination of major portions of the cancer cell genome³². As gatekeeping mutations provide fundamental insights into the biology and pathogenesis of particular cancers and are of singular importance to future diagnostic and therapeutic strategies, further research on this topic should be a priority.

The producers. As noted above, it appears that the cells of solid tumors must accumulate several rate-limiting mutations in cancer

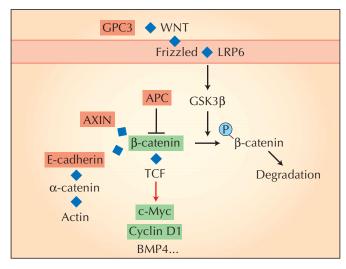


Figure 5 APC pathway. Symbols as in Figure ${\bf 1}$.

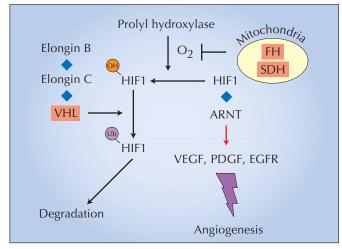


Figure 4 HIF1 pathway. Symbols as in Figure 1.

genes to achieve malignant status. If these mutations had to occur simultaneously in a single cell, then the prevalence of cancer would be minimal³³. The current doctrine is that these mutations occur over time, with each mutation engendering a clonal expansion resulting in a large number of cells that then form a substrate for subsequent mutations⁶. Are normal rates of mutation, coupled with clonal expansions, sufficient to account for the prevalence of cancer, or is some form of genetic instability required for a cell to undergo these multiple, sequential mutations? This issue has been hotly debated for a long time^{34,35}, but the last 10 years of research have yielded some facts that clarify the issues. First, genetic instability can clearly contribute to cancer, as cogently demonstrated by the hereditary cancer predispositions caused by defects in stability genes (Table 1). Second, most cancers do not have a high mutation rate when this rate is measured at the nucleotide level. Accordingly, the number of mutations in a typical cancer cell is ~1 per megabase of DNA, similar to what would be expected in a normal cell that had passed through as many generations and population bottlenecks³⁶. These observations argue against a common role for defects in MMR, NER or BER in nonhereditary tumor types. However, there is another form of instability—chromosomal instability (CIN)—that is much more commonly found in cancers, and this is observed at the gross chromosomal rather than the nucleotide level³⁷. Though the actual rate of chromosomal changes has only been studied in a small number of cases, the end result of CIN, aneuploidy, is observed in nearly all solid tumors³⁸. At the molecular level, chromosome losses are evident as losses of heterozygosity (LOH). An average of 25-30% of the alleles present in normal cells are lost in cancers, and it is not unusual to observe losses of over 75% of the cell's alleles³⁷. Classic as well as modern cytogenetic studies are fully consistent with these observations^{23,39}. Such wholesale changes in chromosomal content can be advantageous for the cancer cell, allowing the efficient elimination of one allele of a tumor-suppressor gene as well as the production of variants that can rapidly adapt to changing microenvironments. The mechanisms underlying aneuploidy and chromosomal instability in sporadic tumors are still largely unknown, though some candidate genes and pathways, such as those involving cell-cycle checkpoints, telomere crisis or centrosomes, have been proposed^{34,40–44}. The role of telomeres in instability is particularly intriguing given the potential importance of telomerase in aging and the agedependent incidence of most cancers 42,45. As genetic instabilities



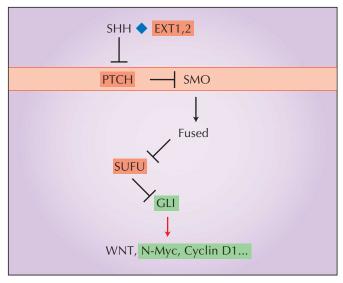


Figure 6 GLI pathway. Symbols as in Figure 1.

not only seem to be central to the neoplastic development but also may underlie the development of resistance to chemotherapeutic agents^{33,34}, the identification of the molecular mechanisms responsible for them is an important area of study.

The ending. Although abnormalities of the cancer genes listed in Tables 1 and 2 are essential contributors to cancer, most abnormalities in these genes occur relatively early in the disease process and none are known to be specifically associated with the metastatic stage. It is this final stage—the seeding and growth of satellite lesions in other organs—that is ultimately responsible for the great majority of neoplastic deaths⁴⁶. Primary tumors can generally be removed through surgery but widely metastatic lesions cannot be excised and are difficult or impossible to treat with adjuvant therapies.

Some of the biochemical processes involved in the early stages of metastasis, such as increased cell motility and production of matrix-degrading proteases, have been well studied⁴⁷. However, the genetic alterations responsible for endowing cells with these abilities have not been clearly identified. In fact, it has been suggested that metastasis is not dependent at all on new genetic abnormalities that occur after tumors have been established, and that the propensity to metastasize is determined early in the neoplastic process rather than near its end (reviewed in ref. 48). This suggestion is not readily compatible with the evidence that cancer is a genetic disease in which evolution occurs somatically. Just as macroevolution never stops, evolution of the cancer cell does not stop and new variants of tumor cells with potentially greater capacities to invade and metastasize are always being born. This evolution is itself driven by inherent genetic instabilities, as described above. However, this genetic perspective on metastasis cannot be validated—or invalidated—until a better understanding of the metastatic process is in hand.

Same actors in different roles. One might have expected that a specific mutation of a widely expressed gene would have identical or at least similar effects in different mammalian cell types. But this is not in general what is observed. Different effects of the same mutation are not only found in distinct cell types; differences can even be observed in the same cell type, depending on when the mutation occurred during the tumorigenic process. RAS gene mutations provide informative examples of these complexities.

i. Cell type specificity: KRAS2 gene mutations in normal pancreatic duct cells seem to initiate the neoplastic process, eventually leading to the development of pancreatic cancer^{49,50}. The same mutations occurring in normal colonic or ovarian epithelial cells lead to self-limiting hyperplastic or borderline lesions that do not progress to malignancy^{51–53}.

ii. Chronology: In contrast to the effects of KRAS2 mutations in a normal colonic epithelial cell, a KRAS2 gene mutation in the same cell type that has already acquired an APC mutation results in a clonal expansion that often progresses to cancer⁵⁴.

iii. Growth inhibition versus growth promotion: In many human and experimental cancers, RAS genes seem to function as oncogenes^{55,56}. But RAS genes can function as suppressor genes under other circumstances, inhibiting tumorigenesis after administration of carcinogens to mice^{57,58}.

These and similar observations on other cancer genes are consistent with the emerging general notion that signaling molecules play multiple roles at multiple times, even in the same cell type (for example, see ref. 59). However, the biochemical bases for such variations among cancer cells are almost entirely unknown. One could argue that oncogenes and tumor-suppressor genes are like electronic components whose effects depend on their placement within an electrical circuit. But no such argument can be easily used to explain the cell type specificity of stability gene defects. For example, MMR genes seem to have the identical, nonredundant function in every cell type on the planet: they limit the acquisition of specific types of mutations, such as those in homopolymer tracts^{60,61}. Yet inherited mismatch repair defects lead to tumors in the colon and endometrium but spare most other organs, including rapidly dividing, selfrenewing tissues such as the small intestine and bone marrow 62-64.

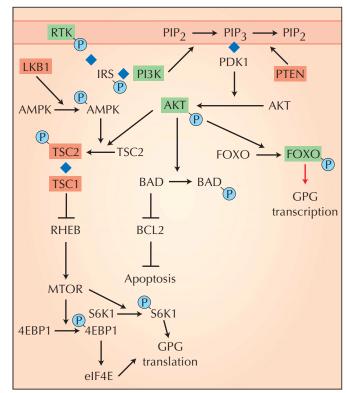


Figure 7 PI3K pathway. Symbols as in Figure 1.

It is also notable in this context that many cancer genes affect different organs when mutated in mice than when mutated in humans⁸. An important practical implication of all this complexity is that it is risky to generalize the conclusions of experiments performed in any cell type or to infer the function of a gene in human cancer cells based on studies of the homologous genes in other organisms.

Major cast members or bit players? In addition to the genes that are mutated in a significant portion of cancers of a given type, such as those listed in Tables 1 and 2, there are many other genes that have been implicated in neoplasia but not shown to be mutated. These genes have been shown to be expressed at higher or lower levels than expected in normal cells^{65,66} and are often associated with 'epigenetic' changes—that is, covalent modifications of DNA or chromatin that are preserved as the cancer cells divide^{67,68}. Unlike genetic changes, epigenetic changes identical to those found in cancers are often found in normal cells at some stage of development.

The discovery of such genes is a growth industry now that high-throughput methods for evaluating the genes expressed in cancer cells have been developed^{65,66,69–74}. The information gained from these studies has proven extremely promising for the development of diagnostic assays, particularly for prognosis (for example, see ref. 75). But as has been shown for prostate-specific antigen (PSA), the utility of a gene for cancer diagnostics does not necessarily reflect a causative role in the process. It will therefore be essential to determine how genes discovered through expression-based approaches can be elevated from candidate status to culprit status in human neoplasia. When genes are sometimes inactivated by mutation and in other cases inactivated by epigenetic silencing, a

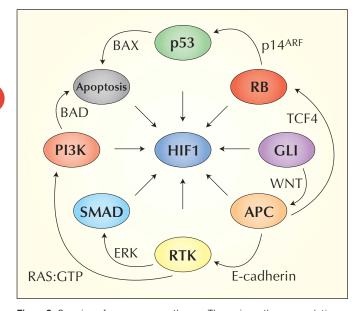


Figure 9 Overview of cancer gene pathways. The major pathways regulating cell birth and cell death are depicted as ovals color-coded to match Figs. 1–8. The schematics in Figs. 1–8 emphasize the genes that have been shown to be genetically altered in human tumors, though many other genes participate in these pathways. Additionally, some of the same genes appear in more than one pathway and there is substantial 'cross-talk' between pathways. Selected mediators of this cross-talk are indicated in the loops that connect the pathways. More detailed information about these pathways can be found in several comprehensive reviews (refs. 11,12,15,31,103–116).

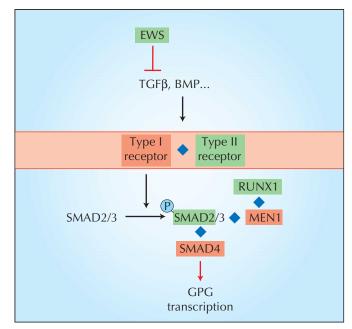


Figure 8 SMAD pathway. Symbols as in Figure 1.

cogent case can be made for their involvement in cancer. Relevant examples include the VHL and CDKN2A tumor-suppressor genes and the hMLH1 stability gene⁶⁷. In the absence of mutational evidence, functional evidence obtained from studies in vitro or in nonhuman species in vivo must be used. But because of the issues with cell type, species and chronological specificity noted above, expression and functional studies cannot currently provide the 'smoking guns' that would definitively implicate the gene in human cancers^{76,77}. Telomerase provides a cogent example of the difficulties involved in such determinations. There is abundant evidence implicating telomerase in processes characteristic of neoplasia, such as immortalization^{45,78} and genetic instability^{42,79}, and a great deal is known about its biochemical properties^{80,81}. Even so, it is unclear at present whether the expression of telomerase in cancers is abnormal, driving the neoplastic process, or simply reflects the fact that cancers are derived from stem cells that normally express this enzyme.

Levels of gene expression are unreliable indicators of causation because disturbance of any network invariably leads to a multitude of such changes only peripherally related to the phenotype⁸². Without better ways to determine whether an unmutated but interesting candidate gene has a causal role in neoplasia, cancer researchers will likely be spending precious time working on genes only peripherally related to the disease they wish to study. One challenge for the future is therefore to develop new model systems. For liquid tumors, innovative systems have indeed been developed, such as those using immunodeficient mice reconstituted with human hematopoietic stem cells⁸³. Perhaps analogous humanized models can be developed to explore solid tumors in a more medically meaningful context than is currently possible^{84,85}.

What makes a box-office smash? Significant gains in cancer therapeutics have also been made in the last decade. Many of these advances have been incremental, but the increments add up. Thus more potent, less toxic derivatives of classic chemotherapeutic agents have been developed, dosing and combination treatments further optimized and side effects ameliorated⁸⁶. However, the



Table 2 Genes that are mutated somatically but not inherited in mutant form

Gene ^a (synonym)	Somatic mutation type ^b	Cancers with mutant gene ^c	Pathway ^d
CTNNB1 (β-catenin)	Activating codon change	Colon, liver, medulloblastomas	APC
BCL2	Translocation	Lymphomas	APOP
TNFRSF6 (FAS)	Activating codon change	Lymphomas, testicular germ cell tumors	APOP
BAX	Inactivating codon change	Colon, stomach	APOP
FBXW7 (CDC4)	Inactivating codon change	Colon, uterine, ovarian, breast	CIN
GLI	Amplification, translocation	Brain, sarcomas	GLI
HPVE6	HPV infection	Cervical	p53
MDM2	Amplification	Sarcomas	p53
NOTCH1	Translocation	Leukemias	p53
AKT2	Amplification	Ovarian, breast	PI3K
FOXO1A, 3A	Translocation	Rhabdomyosarcomas, leukemias	PI3K
PI3KCA	Activating codon change	Colon, stomach, brain, breast	PI3K
CCND1 (cyclin D1)	Amplification, translocation	Leukemias, breast	RB
HPVE7	HPV infection	Cervical	RB
TAL1	Translocation	Leukemias	RB
TFE3	Translocation	Kidney, sarcomas	RB
ABL1 (ABL)	Translocation	Chronic myelogenous leukemia	RTK
ALK	Translocation	Anaplastic large cell lymphoma	RTK
BRAF	Activating codon change	Melanoma, colorectal, thyroid	RTK
EGFR	Amplification, activating codon change	Glioblastomas, non-small cell lung cancers	RTK
EPHB2	Inactivating codon change	Prostate	RTK
ERBB2	Amplification	Breast, ovarian	RTK
FES	Activating codon change	Colon	RTK
FGFR1–3	Translocation	Lymphomas, gastric cancers, bladder cancers	RTK
FLT3, 4	Activating codon change	Leukemias, angiosarcomas	RTK
JAK2	Translocation	Leukemias	RTK
KRAS2, N-RAS	Activating codon change	Colorectal, pancreatic, non-small cell lung cancer	RTK
NTRK1, 3	Translocation, activating codon change	Thyroid, secretory breast, colon	RTK
PDGFB	Translocation	Dermatofibrosarcomas and fibroblastomas	RTK
PDGFRB	Translocation	Leukemias	RTK
EWSR1	Translocation	Ewing's sarcomas, lymphomas, leukemias	SMAD
RUNX1	Translocation	Leukemias	SMAD
SMAD2	Inactivating codon change	Colon, breast	SMAD
TGFBR1, TGFBR2	Inactivating codon change	Colon, stomach, ovarian	SMAD
BCL6	Translocation	Lymphomas	?
EVI1	Translocation	Leukemias	?
HMGA2	Translocation	Lipomas	?
HOXA9, 11, 13; HOXC13,	Translocation	Leukemias	?
HOXD11, 13; HOX11, HOX11L2			
MAP2K4 (MKK4)	Inactivating codon change	Pancreas, breast, colon	?
MLL	Translocation, activating codon change	Leukemias	?
MYC, MYCN, MYCL1	Amplification	Lymphomas, neuroblastomas, small cell lung cancers	?
PTNP1, 11	Activating codon change	Leukemias, colon	?
RARA	Translocation	Promyelocytic leukemia	?
SS18	Translocation	Synovial sarcomas	?

aRepresentative genes of all the major pathways and cancer types are listed. For a complete list, see ref. 76. Approved gene symbols are provided for each entry, with alternative names in parentheses. ^bActivating codon change, intragenic mutation altering one or a small number of base pairs that activates the gene product, indicating that it is an oncogene; inactivating codon change any mutation (point mutation, small or large deletion, etc.) that inactivates the gene product, indicating that the gene is a tumor suppressor. Amplifications and translocations generally affect oncogenes, though occasional translocations disrupt a gene rather than activate it (such as has been suggested to occur with *RUNX1*; ref. 118). ^{(Only representative types of cancers are listed when a gene is mutated in many tumor types, Specific types of leukemias and lymphomas are listed only if a given gene is predominantly mutated in a specific subtype. ^(In) In many cases, the gene has been implicated in several pathways. The single pathway that is listed for each gene represents a 'best guess' (when one can be made) and for the reasons noted in the text and in the legend to Figure 9, should not be regarded as conclusive. APOP, apoptotic pathway; RTK, receptor tyrosine kinase pathway (see Fig. 9).}

most dramatic therapeutic advances have come from agents targeted against proteins encoded by genes that are mutated in cancers. These include trastuzumab (Herceptin), an antibody against the product of the *ERBB2* gene amplified in some breast cancers⁸⁷; imatinib (Gleevec), an inhibitor of tyrosine kinases altered in chronic myelogenous leukemia (CML)⁸⁸ and gastrointestinal stromal tumors (GISTs)⁸⁹; and gefitinib (Iressa), an EGFR

kinase inhibitor used to treat lung cancers⁹⁰. Though none of these advances have resulted in cures of many patients with advanced disease, they can substantially improve and prolong lives. One important lesson from the use of these agents is that mutations are more reliable indicators of a good target than is abnormal expression. For example, virtually all GISTs abnormally express high levels of c-kit, but only those tumors with intragenic mutations of *KIT*



npg © 2004 Natur

respond to therapy with imatinib⁹¹. Similarly, gefitinib is efficacious only in lung cancers in which the kinase domain of EGFR is mutated but not in lung cancers in which EGFR is simply overexpressed^{92,93}.

Interestingly, we still don't know the basis for the selectivity of conventional chemotherapeutic agents. Why cancer cells are more sensitive to antimetabolites, alkylating agents, DNA intercalators and topoisomerase inhibitors than normal replicating cells is enigmatic. The explosion in research on apoptosis in the 1990s has suggested that some tumor cells are less, rather than more, likely to undergo apoptosis after noxious stimuli^{94,95}. Differential apoptotic proclivities therefore don't readily explain therapeutic specificities. This is consistent with studies showing that many drugs used in the clinic to treat solid tumors do not function through the induction of apoptosis at the doses achieved in patients⁹⁶. Even with the newer, targeted agents, the basis for selectivity is not generally clear. The ERBB2 and ABL tyrosine kinases, for example, are expressed in many normal cell types; why should targeting these kinases lead to the demise of cancer cells but not normal cells? Concepts such as 'cellular addiction' offer an imaginative answer to this question, but the molecular basis for the postulated addiction is unknown⁹⁷. Similar arguments could be made for 'untargeted' drugs like Taxol; are cancer cells addicted to microtubules? It is interesting that so much work has been done on how cells become resistant to conventional chemotherapeutic agents and so little on why they are sensitive to begin with. Indeed, some studies indicate that it is not the neoplastic cells, but rather the tumor vasculature, that is the actual target of many conventional chemotherapeutic agents²⁷. Perhaps if we understood the bases for the responsiveness of cancers to these drugs, we could devise better agents to exploit them.

The sequel. There are at least three major challenges that will occupy most cancer researchers' time over the next 10 years. The first is the discovery of new genes that have a causal role in neoplasia, particularly those that initiate and conclude the process. The second is the delineation of the pathways through which these genes act and the basis for the varying actions in specific cell types. The third is the development of new ways to exploit this knowledge for the benefit of patients.

The first two of these challenges are likely to proceed apace. Technologies for gene discovery are rapidly advancing, the human genome will soon be in finished form, and previous analyses of other genes provide well-traveled road maps for investigators to follow once new genes are in hand. In fact, there are likely to be too many genes, too many interacting proteins and too many potential functions to consider, rather than too few. As noted above, cancer biology has not kept up with cancer molecular genetics and new biological systems are needed to separate wheat from chaff.

The third challenge, involving practical benefits to patients, will be much more difficult to meet. There are hardly any road maps to follow. Eradicating hundreds of billions of cancer cells from a human with metastatic disease is a daunting task. Each of these cells has multiple genetic abnormalities and is capable of rapidly evolving variants to combat any therapeutic onslaught^{98,99}. On the other hand, many cancers respond to the somewhat crude weapons that have been developed to date despite their multiple genetic abnormalities. And now that cancer is no longer a 'black box', and is known to be the result of alterations in a limited number of pathways that can in principle be targeted by new generations of drugs, cautious optimism is appropriate.

But will the development of new, more specific therapeutic agents be the best way to minimize cancer morbidity and mortality in the long-term? Most of the increased longevity that Western societies enjoy today has come through better prevention rather than better treatment. It is clear that certain major forms of cancer can be prevented by limiting exposures to carcinogens (such as sunlight and cigarette smoke in skin and lung cancers, respectively). Other forms of cancer can be detected early, thereby limiting morbidity and mortality (for example, tumors of the breast, cervix, prostate and colon). Moreover, it requires 30-40 years for a typical epithelial cell to accumulate the multiple genetic alterations required to progress to metastatic disease¹⁰⁰. This provides a huge window of opportunity to detect tumors at a stage when they are still curable by conventional surgical methods. Perhaps knowledge of the genes altered in cancer will provide the basis for new generations of diagnostic tests, employing target-specific imaging or analysis of body fluid samples ex vivo that will make early detection approaches feasible 101,102. Though less dramatic than cures, prevention and early detection are perhaps the most promising and feasible means to reduce cancer deaths by the time that Nature Medicine celebrates its 20th anniversary.

ACKNOWLEDGMENTS

The authors thank many colleagues for their critical reading of the manuscript and S. Mousses for sharing results on EphB2 before publication. Work in the authors' laboratories has been supported by the Ludwig Trust, the National Colorectal Cancer Research Alliance, the Miracle Foundation, the Clayton Fund and US National Institutes of Health grants CA43460, CA57345 and CA62924.

COMPETING INTERESTS STATEMENT

The authors declare competing financial interests (see the *Nature Medicine* website for details).

This material is part of Nature Medicine's 10 year anniversary series. For more content related to these special focus issues, please see http://www.nature.com/nm/special_focus/anniversary/index.html

Published online at http://www.nature.com/naturemedicine/

- Davies, H. et al. Mutations of the BRAF gene in human cancer. Nature 417, 949–954 (2002).
- Wan, P.T. et al. Mechanism of activation of the RAF-ERK signaling pathway by oncogenic mutations of B-RAF. Cell 116, 855–867 (2004).
- Santarosa, M. & Ashworth, A. Haploinsufficiency for tumour suppressor genes: when you don't need to go all the way. *Biochim. Biophys. Acta* 1654, 105–122 (2004).
- 4. Knudson, A.G. Cancer genetics. Am. J. Med. Genet. 111, 96-102 (2002).
- 5. Friedberg, E.C. DNA damage and repair. *Nature* **421**, 436–440 (2003).
- Nowell, P.C. Tumor progression: a brief historical perspective. Semin. Cancer Biol. 12, 261–266 (2002).
- Maley, C.C. et al. Selectively advantageous mutations and hitchhikers in neoplasms: p16 lesions are selected in Barrett's esophagus. Cancer Res. 64, 3414–3427 (2004).
- Van Dyke, T. & Jacks, T. Cancer modeling in the modern era: progress and challenges. *Cell* 108, 135–144 (2002).
- 9. Horvitz, H.R. Worms, life, and death. Chembiochem 4, 697–711 (2003).
- 10. Sherr, C.J. Cancer cell cycles revisited. Cancer Res. 60, 3689–3695 (2000).
- Ortega, S., Malumbres, M. & Barbacid, M. Cyclin D-dependent kinases, INK4 inhibitors and cancer. *Biochim. Biophys. Acta* 1602, 73–87 (2002).
- Classon, M. & Harlow, E. The retinoblastoma tumour suppressor in development and cancer. Nat. Rev. Cancer 2, 910–917 (2002).
- Ichimura, K. et al. Deregulation of the p14ARF/MDM2/p53 pathway is a prerequisite for human astrocytic gliomas with G1-S transition control gene abnormalities. Cancer Res. 60, 417–424 (2000).
- Vogelstein, B., Lane, D. & Levine, A.J. Surfing the p53 network. *Nature* 408, 307–310 (2000).
- Oren, M. Decision making by p53: life, death and cancer. Cell Death Differ. 10, 431–442 (2003).
- 16. Prives, C. & Hall, P.A. The p53 pathway. J. Pathol. 187, 112-126 (1999).
- Klein, G. Perspectives in studies of human tumor viruses. Front. Biosci. 7, d268–d274 (2002).
- Munger, K. & Howley, P.M. Human papillomavirus immortalization and transformation functions. Virus Res. 89, 213–228 (2002).
- 19. zur Hausen, H. Oncogenic DNA viruses. *Oncogene* **20**, 7820–7823 (2001).

- 20. Hunter, T. Signaling-2000 and beyond. Cell 100, 113-127 (2000).
- Komarova, N.L., Sengupta, A. & Nowak, M.A. Mutation-selection networks of cancer initiation: tumor suppressor genes and chromosomal instability. *J. Theor. Biol.* 223, 433–450 (2003).
- 22. Rowley, J.D. The critical role of chromosome translocations in human leukemias. *Annu. Rev. Genet.* **32**, 495–519 (1998).
- Mitelman, F. Recurrent chromosome aberrations in cancer. Mutat. Res. 462, 247–253 (2000).
- Verheul, H.M., Voest, E.E. & Schlingemann, R.O. Are tumours angiogenesisdependent? J. Pathol. 202, 5–13 (2004).
- Tlsty, T.D. & Hein, P.W. Know thy neighbor: stromal cells can contribute oncogenic signals. Curr. Opin. Genet. Dev. 11, 54–59 (2001).
- Fata, J.E., Werb, Z. & Bissell, M.J. Regulation of mammary gland branching morphogenesis by the extracellular matrix and its remodeling enzymes. *Breast Cancer Res.* 6, 1–11 (2004).
- 27. Kerbel, R. & Folkman, J. Clinical translation of angiogenesis inhibitors. *Nat. Rev. Cancer* **2**, 727–739 (2002).
- Folkman, J. Role of angiogenesis in tumor growth and metastasis. Semin. Oncol. 29, 15–18 (2002).
- Ferrara, N., Hillan, K.J., Gerber, H.P. & Novotny, W. Discovery and development of bevacizumab, an anti-VEGF antibody for treating cancer. *Nat. Rev. Drug Discov.* 3, 391–400 (2004).
- Kondo, K., Klco, J., Nakamura, E., Lechpammer, M. & Kaelin, W.G. Jr. Inhibition of HIF is necessary for tumor suppression by the von Hippel-Lindau protein. *Cancer Cell* 1, 237–246 (2002).
- Semenza, G.L. Targeting HIF-1 for cancer therapy. Nat. Rev. Cancer 3, 721–732 (2003).
- Strausberg, R.L., Simpson, A.J. & Wooster, R. Sequence-based cancer genomics: progress, lessons and opportunities. *Nat. Rev. Genet.* 4, 409–418 (2003).
- Loeb, L.A., Loeb, K.R. & Anderson, J.P. Multiple mutations and cancer. Proc. Natl. Acad. Sci. USA 100, 776–781 (2003).
- Rajagopalan, H., Nowak, M.A., Vogelstein, B. & Lengauer, C. The significance of unstable chromosomes in colorectal cancer. *Nat. Rev. Cancer* 3, 695–701 (2003).
- 35. Sieber, O.M., Heinimann, K. & Tomlinson, I.P. Genomic instability—the engine of tumorigenesis? *Nat. Rev. Cancer* **3**, 701–708 (2003).
- Wang, T.L. et al. Prevalence of somatic alterations in the colorectal cancer cell genome. Proc. Natl. Acad. Sci. USA 99, 3076–3080 (2002).
- Lengauer, C., Kinzler, K.W. & Vogelstein, B. Genetic instabilities in human cancers. Nature 396, 643–649 (1998).
- Duesberg, P. & Li, R. Multistep carcinogenesis: a chain reaction of aneuploidizations. Cell Cycle 2, 202–210 (2003).
- Albertson, D.G. & Pinkel, D. Genomic microarrays in human genetic disease and cancer. Hum. Mol. Genet. 12 (spec. no. 2), R145–R152 (2003).
- Shiloh, Y. & Kastan, M.B. ATM: genome stability, neuronal development, and cancer cross paths. Adv. Cancer Res. 83, 209–254 (2001).
- Scully, R. & Livingston, D.M. In search of the tumour-suppressor functions of BRCA1 and BRCA2. Nature 408, 429–432 (2000).
- Maser, R.S. & DePinho, R.A. Connecting chromosomes, crisis, and cancer. Science 297, 565–569 (2002).
- Pihan, G. & Doxsey, S.J. Mutations and aneuploidy: co-conspirators in cancer? Cancer Cell 4, 89–94 (2003).
- Rajagopalan, H. et al. Inactivation of hCDC4 can cause chromosomal instability. Nature 428, 77–81 (2004).
- Shay, J.W. & Roninson, I.B. Hallmarks of senescence in carcinogenesis and cancer therapy. *Oncogene* 23, 2919–2933 (2004).
- Chambers, A.F., Groom, A.C. & MacDonald, I.C. Dissemination and growth of cancer cells in metastatic sites. *Nat. Rev. Cancer* 2, 563–572 (2002).
- Fidler, I.J. Critical determinants of metastasis. Semin. Cancer Biol. 12, 89–96 (2002).
- 48. Hunter, K.W. Host genetics and tumour metastasis. *Br. J. Cancer* **90**, 752–755 (2004).
- Hruban, R.H., Goggins, M., Parsons, J. & Kern, S.E. Progression model for pancreatic cancer. Clin. Cancer Res. 6, 2969–2972 (2000).
- Aguirre, A.J. et al. Activated Kras and Ink4a/Arf deficiency cooperate to produce metastatic pancreatic ductal adenocarcinoma. Genes Dev. 17, 3112–3126 (2003).
- Jen, J. et al. Molecular determinants of dysplasia in colorectal lesions. Cancer Res. 54, 5523–5526 (1994).
- Pretlow, T.P. Aberrant crypt foci and K-ras mutations: earliest recognized players or innocent bystanders in colon carcinogenesis? *Gastroenterology* 108, 600–603 (1995).
- Sieben, N.L. et al. In ovarian neoplasms, BRAF, but not KRAS, mutations are restricted to low-grade serous tumours. J. Pathol. 202, 336–340 (2004).
- Kinzler, K.W. & Vogelstein, B. Colorectal Tumors. in *The Genetic Basis of Human Cancer* (eds. Vogelstein, B. & Kinzler, K.W.) 565–587 (McGraw-Hill, New York, 1998).
- 55. Barbacid, M. ras genes. Annu. Rev. Biochem. 56, 779-827 (1987).
- Bos, J.L. ras oncogenes in human cancer: a review. *Cancer Res.* 49, 4682–4689 (1989).
- Zhang, Z. et al. Wildtype Kras2 can inhibit lung carcinogenesis in mice. Nat. Genet. 29, 25–33 (2001).
- 58. Diaz, R. et al. The N-ras proto-oncogene can suppress the malignant phenotype

- in the presence or absence of its oncogene. Cancer Res. 62, 4514-4518 (2002).
- Bronner-Fraser, M. Development. Making sense of the sensory lineage. Science 303, 966–968 (2004).
- Jiricny, J. Eukaryotic mismatch repair: an update. Mutat. Res. 409, 107–121 (1998).
- Fishel, R. & Wilson, T. MutS homologs in mammalian cells. Curr. Opin. Genet. Dev. 7, 105–113 (1997).
- 62. Lynch, H.T. & de la Chapelle, A. Hereditary colorectal cancer. N. Engl. J. Med. 348, 919–932 (2003).
- Yamamoto, H., Imai, K. & Perucho, M. Gastrointestinal cancer of the microsatellite mutator phenotype pathway. J. Gastroenterol. 37, 153–163 (2002)
- 64. Honchel, R., Halling, K.C. & Thibodeau, S.N. Genomic instability in neoplasia. Semin. Cell Biol. 6, 45–52 (1995).
- Brown, P.O. & Botstein, D. Exploring the new world of the genome with DNA microarrays. Nat. Genet. 21, 33–37 (1999).
- Polyak, K. & Riggins, G.J. Gene discovery using the serial analysis of gene expression technique: implications for cancer research. J. Clin. Oncol. 19, 2948–2958 (2001).
- 67. Jones, P.A. & Baylin, S.B. The fundamental role of epigenetic events in cancer. Nat. Rev. Genet. 3, 415–428 (2002).
- Feinberg, A.P. & Tycko, B. The history of cancer epigenetics. *Nat. Rev. Cancer* 4, 143–153 (2004).
- Collins, F.S., Green, E.D., Guttmacher, A.E. & Guyer, M.S. A vision for the future of genomics research. *Nature* 422, 835–847 (2003).
- Schadt, E.E., Monks, S.A. & Friend, S.H. A new paradigm for drug discovery: integrating clinical, genetic, genomic and molecular phenotype data to identify drug targets. *Biochem. Soc. Trans.* 31, 437–443 (2003).
- Paddison, P.J. et al. A resource for large-scale RNA-interference-based screens in mammals. Nature 428, 427–431 (2004).
- Berns, K. et al. A large-scale RNAi screen in human cells identifies new components of the p53 pathway. Nature 428, 431–437 (2004).
- Rosenblatt, K.P. et al. Serum proteomics in cancer diagnosis and management. Annu. Rev. Med. 55, 97–112 (2004).
- Luo, J., Isaacs, W.B., Trent, J.M. & Duggan, D.J. Looking beyond morphology: cancer gene expression profiling using DNA microarrays. *Cancer Invest.* 21, 937–949 (2003).
- Ma, X.J. et al. A two-gene expression ratio predicts clinical outcome in breast cancer patients treated with tamoxifen. Cancer Cell 5, 607–616 (2004).
- Futreal, P.A. et al. A census of human cancer genes. Nat. Rev. Cancer 4, 177–183 (2004).
- Masayesva, B.G. et al. Gene expression alterations over large chromosomal regions in cancers include multiple genes unrelated to malignant progression. Proc. Natl. Acad. Sci. USA 101, 8715–8720 (2004).
- Stewart, S.A. & Weinberg, R.A. Senescence: does it all happen at the ends? Oncogene 21, 627–630 (2002).
- Feldser, D.M., Hackett, J.A. & Greider, C.W. Telomere dysfunction and the initiation of genome instability. *Nat. Rev. Cancer* 3, 623–627 (2003).
- Chan, S.R. & Blackburn, E.H. Telomeres and telomerase. *Phil. Trans. R. Soc. Lond. B* 359, 109–121 (2004).
- Cech, T.R. Beginning to understand the end of the chromosome. Cell 116, 273–279 (2004).
- Miklos, G.L. & Maleszka, R. Microarray reality checks in the context of a complex disease. Nat. Biotechnol. 22, 615–621 (2004).
- Hope, K.J., Jin, L. & Dick, J.E. Human acute myeloid leukemia stem cells. Arch. Med. Res. 34, 507–514 (2003).
- Berking, C. & Herlyn, M. Human skin reconstruct models: a new application for studies of melanocyte and melanoma biology. *Histol. Histopathol.* 16, 669–674 (2001).
- Kuperwasser, C. et al. Reconstruction of functionally normal and malignant human breast tissues in mice. Proc. Natl Acad. Sci. USA 101, 4966–4971 (2004).
- Frei, E.I. & Eder, J.P. Principles of dose, schedule, and combination Therapy. in *Cancer Medicine* (eds. Kufe, D.W. et al.) 669–677 (B.C. Decker, Inc., Hamilton, Ontario, 2003).
- Pegram, M.D., Konecny, G. & Slamon, D.J. The molecular and cellular biology of HER2/neu gene amplification/overexpression and the clinical development of herceptin (trastuzumab) therapy for breast cancer. *Cancer Treat. Res.* 103, 57–75 (2000).
- Druker, B.J. et al. Chronic myelogenous leukemia. in Hematology 2001 (American Society of Hematology Education Program) 87–112 (American Society of Hematology, 2001).
- Mechtersheimer, G. et al. Gastrointestinal stromal tumours and their response to treatment with the tyrosine kinase inhibitor imatinib. Virchows Arch. 444, 108–118 (2004).
- Langer, C.J. Emerging role of epidermal growth factor receptor inhibition in therapy for advanced malignancy: focus on NSCLC. *Int. J. Radiat. Oncol. Biol. Phys.* 58, 991–1002 (2004).
- Duensing, A., Heinrich, M.C., Fletcher, C.D. & Fletcher, J.A. Biology of gastrointestinal stromal tumors: KIT mutations and beyond. *Cancer Invest.* 22, 106–116 (2004).
- Paez, J.G. et al. EGFR mutations in lung cancer: correlation with clinical response to gefitinib therapy. Science 304, 1497–1500 (2004).



- 93. Lynch, T.J. *et al.* Activating mutations in the epidermal growth factor receptor underlying responsiveness of non-small-cell lung cancer to gefitinib. *N. Engl. J. Med.* **350**, 2129–2139 (2004).
- Schmitt, C.A. & Lowe, S.W. Apoptosis and therapy. J. Pathol. 187, 127–137 (1999).
- Danial, N.N. & Korsmeyer, S.J. Cell death: critical control points. Cell 116, 205–219 (2004).
- Brown, J.M. & Wouters, B.G. Apoptosis: mediator or mode of cell killing by anticancer agents? *Drug Resist. Updat.* 4, 135–136 (2001).
- Weinstein, I.B. et al. Disorders in cell circuitry associated with multistage carcinogenesis: exploitable targets for cancer prevention and therapy. Clin. Cancer Res. 3, 2696–2702 (1997).
- Nygren, P. & Larsson, R. Overview of the clinical efficacy of investigational anticancer drugs. J. Intern. Med. 253, 46–75 (2003).
- Shih, L.Y. et al. Heterogeneous patterns of FLT3 Asp(835) mutations in relapsed de novo acute myeloid leukemia: a comparative analysis of 120 paired diagnostic and relapse bone marrow samples. Clin. Cancer Res. 10, 1326–1332 (2004).
- Kinzler, K.W. & Vogelstein, B. Lessons from hereditary colon cancer. Cell 87, 159–170 (1996).
- Weissleder, R. & Ntziachristos, V. Shedding light onto live molecular targets. Nat. Med. 9, 123–128 (2003).
- 102. Sidransky, D. Emerging molecular markers of cancer. *Nat. Rev. Cancer* 2, 210–219 (2002).
- 103. Gschwind, A., Fischer, O.M. & Ullrich, A. The discovery of receptor tyrosine kinases: targets for cancer therapy. *Nat. Rev. Cancer* **4**, 361–370 (2004).
- 104. Downward, J. Targeting RAS signalling pathways in cancer therapy. *Nat. Rev. Cancer* **3**, 11–22 (2003).
- 105. Malumbres, M. & Barbacid, M. To cycle or not to cycle: a critical decision in

- cancer. Nat. Rev. Cancer 1, 222-231 (2001).
- Giles, R.H., van Es, J.H. & Clevers, H. Caught up in a Wnt storm: Wnt signaling in cancer. *Biochim. Biophys. Acta* 1653, 1–24 (2003).
- 107. Cantley, L.C. The phosphoinositide 3-kinase pathway. *Science* **296**, 1655–1657 (2002).
- 108. Shi, Y. & Massague, J. Mechanisms of TGF- β signaling from cell membrane to the nucleus. *Cell* **113**, 685–700 (2003).
- Ruiz i Altaba. A., Stecca, B. & Sanchez, P. Hedgehog-Gli signaling in brain tumors: stem cells and paradevelopmental programs in cancer. *Cancer Lett.* 204, 145–157 (2004).
- 110. Adams, J.M. Ways of dying: multiple pathways to apoptosis. *Genes Dev.* 17, 2481–2495 (2003).
- 111. Blagosklonny, M.V. & Pardee, A.B. The restriction point of the cell cycle. *Cell Cycle* 1, 103–110 (2002).
- 112. Plas, D.R. & Thompson, C.B. Cell metabolism in the regulation of programmed cell death. *Trends Endocrinol. Metab.* **13**, 75–78 (2002).
- 113. Green, D.R. & Evan, G.I. A matter of life and death. Cancer Cell 1, 19–30 (2002).
- 114. Eng, C., Kiuru, M., Fernandez, M.J. & Aaltonen, L.A. A role for mitochondrial enzymes in inherited neoplasia and beyond. *Nat. Rev. Cancer* 3, 193–202 (2003).
- 115. Lum, L. & Beachy, P.A. The Hedgehog response network: sensors, switches, and routers. *Science* **304**, 1755–1759 (2004).
- Brivanlou, A.H. & Darnell, J.E. Jr. Signal transduction and the control of gene expression. Science 295, 813–818 (2002).
- 117. Vogelstein, B. & Kinzler, K.W. *The Genetic Basis of Human Cancer* (McGraw-Hill, Toronto, 2002).
- 118. Cameron, E.R. & Neil, J.C. The Runx genes: lineage-specific oncogenes and tumor suppressors. *Oncogene* 23, 4308–4314 (2004).

